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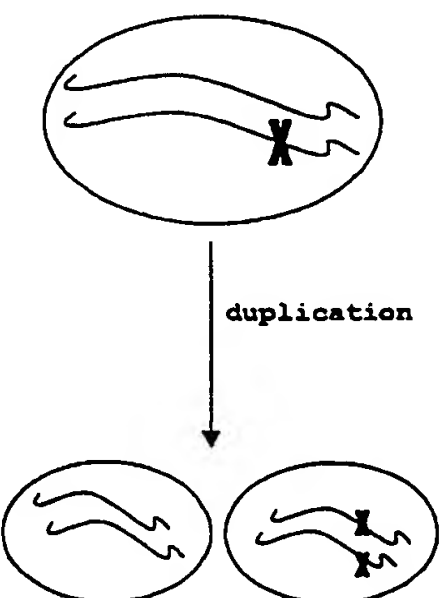
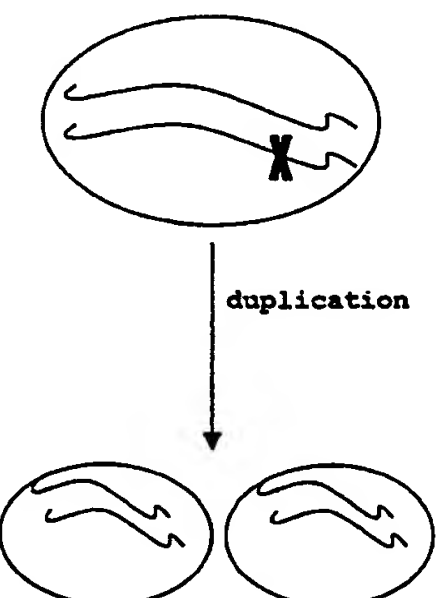
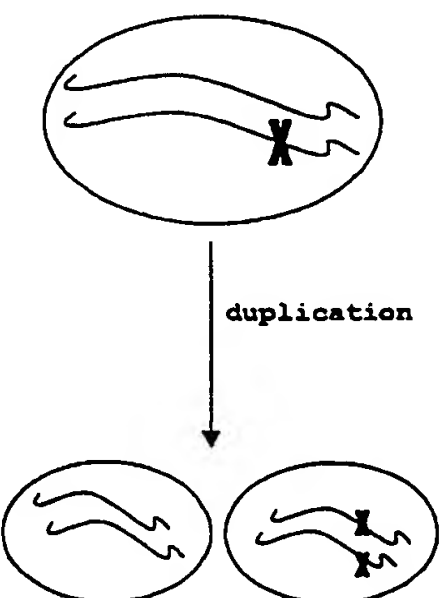
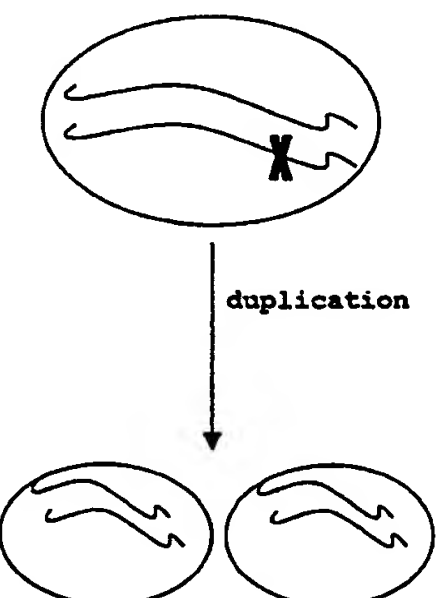
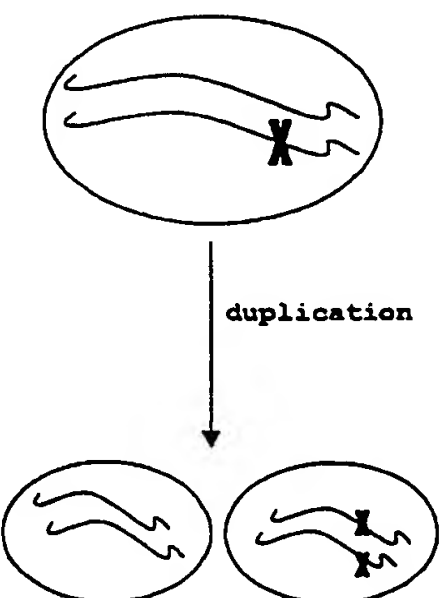
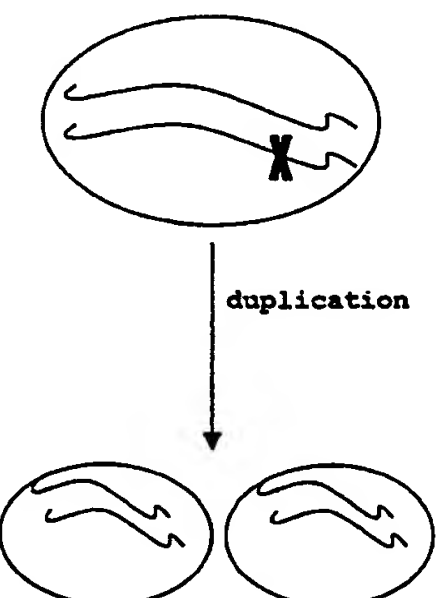
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(54) Title: FUNGAL CELLS WITH INACTIVATED DNA MISMATCH REPAIR SYSTEM <table border="0" style="width: 100%;"><tr><td style="text-align: center; width: 50%;">Mismatch repair <u>inactive</u> ↓  Diversity: <u>Two</u></td><td style="text-align: center; width: 50%;">Mismatch repair <u>active</u> ↓  Diversity: <u>One</u></td></tr></table>			Mismatch repair <u>inactive</u> ↓  Diversity: <u>Two</u>	Mismatch repair <u>active</u> ↓  Diversity: <u>One</u>
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(57) Abstract A process for making DNA libraries in filamentous fungal cells using a novel cloned gene involved in the mismatch repair system of filamentous fungal cells. <div style="text-align: right;">P DOCKET.NO.3123-4006 Ref. to SN 09/834,434</div>				

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FUNGAL CELLS WITH INACTIVATED DNA MISMATCH REPAIR SYSTEM

5 FIELD OF INVENTION

A process for making DNA libraries in filamentous fungal cells using a novel cloned gene involved in the mismatch repair system of filamentous fungal cells.

10 BACKGROUND OF THE INVENTION

The mismatch repair system is a system within cells which recognises mismatches in newly synthesised duplex DNA sequences.

The mismatch repair system then either corrects the
15 mismatches which are seen as errors by e.g. using the methylated ``old'' strain as template or alternatively it may mediate degradation of the duplex DNA sequences which comprise the mismatches.

Independently on the precise mechanism the end result
20 will be that the ``mismatch repair system'' will limit the ``diversity'' within a cell, diversity being represented as duplex DNA sequences which comprise mismatches.

For example a duplex DNA sequence which comprises a single mismatch represents a diversity of two different DNA
25 sequences within the cell. If the mismatch repair system corrects the mismatch there will only be a diversity of one within the cell.

Alternatively, if the mismatch repair system mediates the degradation of such a duplex DNA sequence the diversity will be
30 lost. See figure 1 for a graphic illustration on how the mismatch repair system may work within a cell.

Consequently, if duplex DNA sequences comprising mismatches represent a DNA library of interest, then the diversity of this library may be limited when transformed
35 (placed) into cells with an active mismatch repair system.

The art provides a solution to this problem by making cells wherein the mismatch system is inactive.

EP 449923 describes bacterial cells wherein the mismatch system is inactivated.

WO 97/37011 describes yeast cells wherein the mismatch system is inactivated. See the working examples of this
5 document.

WO 97/05268 describes mice cells wherein the mismatch system is inactivated. See the working examples of this document.

10 SUMMARY OF THE INVENTION:

The problem to be solved by the present invention is to provide an improved strategy for making DNA libraries in filamentous fungal cells. A filamentous fungal cell population comprising such a DNA library may then be used to select a
15 polypeptide of interest. Also polynucleotide sequences with particular properties might be selected, such as promoters, terminators and other regulatory elements with changed/improved properties.

The solution is based on that the present inventors have
20 cloned a NOVEL gene involved in the mismatch repair system of a filamentous fungal cell. Further, this gene is the first gene cloned which is involved in the mismatch repair system of a filamentous fungal cell.

By inactivating this gene in a filamentous cell it is
25 possible to obtain a filamentous cell which is deficient in its mismatch repair system and which is highly useful for preparing DNA libraries in filamentous fungal cells.

The gene comprises a very characterising DNA sequence encoding the polypeptide sequence shown in positions 683-758 of
30 SEQ ID NO 2.

This DNA has been used to clone the full length gene encoding the polypeptide sequence shown in positions 1-940 of SEQ ID NO 2. See working examples herein (vide infra).

The gene cloned as described in working examples herein
35 is a gene cloned from an *Aspergillus oryzae* filamentous fungal cell.

However, based on the novel sequence information provided herein it is routine work for the skilled person to clone

similar homologous genes from other filamentous fungal cells by e.g. standard hybridisation or PCR technology, preferably by using the DNA sequence encoding the polypeptide sequence shown in positions 683-758 of SEQ ID NO 2 as a basis for making a hybridisation probe or PCR primers.

Accordingly, in a first aspect the present invention relates to a filamentous fungal cell, wherein a gene involved in the mismatch repair system has been inactivated and in which the gene involved in the mismatch repair system comprises:

- (a) a DNA sequence encoding the polypeptide sequence shown in positions 683-758 of SEQ ID NO 2; or
- (b) a DNA sequence encoding a polypeptide sequence which is at least 70% identical to the polypeptide sequence shown in positions 683-758 of SEQ ID NO 2;

and

in a second aspect the present invention relates to a filamentous fungal cell, wherein a gene involved in the mismatch repair system has been inactivated and in which the gene involved in the mismatch repair system comprises:

- (a) a DNA sequence encoding the polypeptide sequence shown in positions 1-940 of SEQ ID NO 2; or
- (b) a DNA sequence encoding a polypeptide sequence which is at least 70% identical to the polypeptide sequence shown in positions 1-940 of SEQ ID NO 2.

As stated above a filamentous fungal cell of the first or second aspect of the invention is very suitable for making a DNA library of interest in filamentous fungal cells.

Accordingly, in a third aspect the present invention relates to a process for preparing a filamentous fungal cell population wherein individual cells in the population comprise individually different DNA sequences of interest representing a DNA library of interest comprising following steps:

- (a) placing individually different DNA sequences of interest in a filamentous fungal cell population comprising a filamentous fungal cell of the first or second aspect of the invention; and
- (b) growing the population of (a) for a period of time allowing an individual DNA sequence of interest in the

population to be duplicated at least once under conditions wherein the mismatch repair system gene of the first or second aspect of the invention has been inactivated.

5

One of the advantages of allowing an individual mismatch repair inactivated filamentous fungal cell duplicated DNA of interest at least once as described under step (b) of the third aspect is illustrated in figure 1. As can be seen in figure 1
10 the process of the third aspect using a filamentous fungal mismatch repair inactivated cell as described herein allows preparation of a DNA library wherein eventual hetero duplex DNA mismatches are not corrected. This gives a DNA library with a higher diversity as compared to a DNA library made in a filamentous fungal cell NOT having an inactivated mismatch repair
15 system (see figure 1). Duplication of DNA sequence of interest means that the two strands are replicated such that two separate sets of double stranded DNA are generated, each being based on a separate one of the two original strands.

20 A filamentous fungal cell population wherein individual cells in the population comprise a DNA library of interest as described above may be used to select a polypeptide of interest.

Accordingly, in a fourth aspect the present invention
25 relates to a process for production of a polypeptide of interest comprising the steps of the third aspect and wherein the DNA sequences of interest encode a polypeptide of interest and which further comprises following step:

(c) selecting from the resultant population of
30 filamentous fungal cells of step (b) of the third aspect a desired polypeptide of interest.

An advantage of the process of the fourth aspect may be that the polypeptide of interest is selected from a filamentous fungal cell expressing the polypeptide. Consequently, it is
35 directly known that the polypeptide can be expressed from a filamentous fungal cell, which is usefully if it is subsequently required to produce the polypeptide in large scale in a filamentous fungal cell. This may be of particular

interest when the DNA library encodes polypeptides of interest which are derived from filamentous fungal cells, since it is known that filamentous fungal polypeptides preferably are produced in industrial relevant high yields in filamentous
5 fungal cells.

This is contrary to a similar selection process using e.g. a yeast cell. Here the only thing known is that the selected polypeptide is capable of being expressed in yeast and later expression a filamentous fungal cell might give problems,
10 especially if high yields are required.

DEFINITIONS:

Following section provides definitions of technical features in above-mentioned aspects of the invention.

15 The term "a gene" denotes herein a gene (a DNA sequence) will is capable of being expressed into a polypeptide within said cell. Accordingly, said gene sequence will be defined as an open reading frame starting from a start codon (normally "ATG", "GTG", or "TTG") and ending at a stop codon
20 (normally "TAA", TAG" or "TGA").

In order to express said gene there must be elements, as known in the art, in connection with the gene, necessary for expression of the gene within the cell. Such standard elements may include a promoter, a ribosomal binding site, a termination
25 sequence, and may be others elements as known in the art.

The term "mismatch repair system" shall herein be understood according to the art, as a system within cells which recognises mismatches in duplex DNA sequences. See e.g. WO 97/37011, page 1, line 21-28)

30 The mismatch repair system then either corrects the mismatches which are seen as errors by e.g. using the methylated "old" strain as template or alternatively it may mediate degradation of the duplex DNA sequences which comprise the mismatches.

35 Independently on the precise mechanism the end result will be that the "mismatch repair system" will limit the "diversity" within the cell represented by such duplex DNA sequences which comprise mismatches.

For example a duplex DNA sequence which comprises a single mismatch represents a diversity of two different DNA sequences within the cell. If the mismatch repair system corrects the mismatch there will only be a diversity of one 5 within the cell. Alternatively, if the mismatch repair system mediates the degradation of such a duplex DNA sequence this diversity will be lost.

A polypeptide encoded by a gene involved in the mismatch repair system recognises a mismatch by a mechanism involving 10 binding to the mismatch.

Accordingly, a suitable assay to test whether or not a filamentous fungal cell as described herein is inactivated in its mismatch repair system is to use a "gel shift assay" or alternatively termed a "gel retardation assay". This is a 15 standard assay used in the art. See WO 97/05268, page 16,17 and 25.

The principle in such an assay is that cell extracts are prepared of both (a) a filamentous fungal cell wherein the gene, as described herein, involved in the mismatch repair 20 system is inactivated; and (b) the corresponding filamentous fungal cell wherein the gene is NOT inactivated. These extracts are then bound/mixed to oligonucleotides containing the base-pair mismatched G:T; G:A; G:G; A:C, and an extrahelical TG dinucleotide and run on a nondenaturing gel.

25 If the gel shift assay demonstrates that the control filamentous fungal cell wherein the gene is NOT inactivated comprises any protein(s) which binds to any of above mentioned oligonucleotides and these binding protein(s) are NOT seen in the filamentous fungal cell wherein the gene, as described 30 herein, involved in the mismatch repair system is inactivated then it is a confirmation that the mismatch repair system in the latter is inactivated.

A detailed description of a suitable gel shift assay is provided in working example 1 herein.

35 The sequence identity in relation to the terms

"a DNA sequence encoding a polypeptide sequence which is at least 70% identical to the polypeptide sequence shown in positions 683-758 of SEQ ID NO 2" and

"a DNA sequence encoding a polypeptide sequence which is at least 70% identical to the polypeptide sequence shown in positions 1-940 of SEQ ID NO 2";

is determined as the degree of identity between two sequences indicating a derivation of the first sequence from the second. The identity may suitably be determined by means of computer programs known in the art such as GAP provided in the GCG program package (Program Manual for the Wisconsin Package, Version 8, August 1994, Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA 53711) (Needleman, S.B. and Wunsch, C.D., (1970), Journal of Molecular Biology, 48, 443-453). Using GAP with the following settings for polypeptide sequence comparison: GAP creation penalty of 3.0 and GAP extension penalty of 0.1, the polypeptide encoded by an analogous DNA sequence of the invention exhibits a degree of identity preferably of at least 70%, more preferably at least 80%, more preferably at least 90%, more preferably at least 95%, and especially at least 97% with amino acid sequence shown in positions 683-758 of SEQ ID NO 2, according to the first aspect of the invention; or with amino acid sequence shown in positions 1-940 of SEQ ID NO 2, according to the second aspect of the invention.

The term "DNA library" denotes herein a library of at least two different DNA sequences. For many practical purposes the library is must bigger. Accordingly, the DNA library preferably comprises at least 1000 different DNA sequences, more preferably at least 10000 different DNA sequences, and even more preferably at least 100000 different DNA sequences.

The term "placing individually different DNA sequences of interest in a filamentous fungal cell population" in relation to step (a) in the process of the third aspect of the invention shall herein be understood broadly in the sense that it is NOT identical DNA sequences of interest which are placed in the filamentous fungal cell population. In the present context, relating to a process for making a DNA library using a mismatch repair deficient cell, the term should preferably denotes a situation wherein a cell within the filamentous fungal cell population comprises at least two different DNA sequences of

interest which are so partially homologous that they are capable of hybridising/recombining to each other within the cell. It is within the skilled persons general knowledge to determine how partially homologous such sequences have to be in order to obtain said recombination within the cell.

A practical example may be that single stranded oligonucleotide sequences partially homologous to chromosomal DNA sequence are placed within the cell or duplex DNA sequences comprising mismatches (e.g. comprised within a vector) are placed within the cell. See below for further description of such examples.

The specific experimental way of placing these DNA sequences within a filamentous cell may be done according to any of the many suitable techniques, such as transformation techniques.

The term "growing the population of (a) for a period of time allowing an individual DNA sequence of interest in the population to be duplicated at least once under conditions wherein the mismatch repair system gene has been inactivated" according to step (b) of the third aspect of the invention denotes that after an individual cell has duplicated itself at least once then the mismatch repair system may be activated again without losing the advantage of the process. The technical reason for this is illustrated in figure 1. In this example a duplex DNA sequence comprising a single mismatch is placed in filamentous cell. After the cell has been duplicated once under conditions wherein the mismatch repair system gene has been inactivated the two individually different single stranded DNA sequences within the duplex DNA have individually been duplicated providing two different duplex sequences, one in each duplicated cell, without any mismatches. Accordingly, since such a cell does NOT comprise duplex DNA sequences of interest having mismatches then there is no technical need to maintaining the mismatch repair system inactivated.

In sections below are described preferred embodiments of the invention by way of examples only.

DRAWINGS:

Figure 1:

This figure illustrates an example wherein a duplex DNA sequence comprising a single mismatch is placed in filamentous cell. After the cell has been duplicated once under conditions wherein the mismatch repair system gene has been inactivated the two individually different single stranded DNA sequences within the duplex DNA have individually been duplicated providing two different duplex sequences, one in each duplicated cell, without any mismatches. On the contrary, in a cell wherein the mismatch repair system is active, a mismatch within a duplex is corrected.

Figure 2:

This figure shows three partial *Aspergillus oryzae* polypeptide sequences: 'msh2'Ao-col10/13/15; derived from cloned PCR fragments. The three partial polypeptide sequences are aligned with two other partial polypeptide sequences of known mismatch repair proteins: a human mismatch repair protein, msh2-human.p; and a fungal *Saccharomyces cerevisiae* mismatch repair protein, S.c. msh2. The underlined sequences in the figure derive from the construction of the PCR fragments.

Figure 3:

This figure shows an alignment of the proposed polypeptide sequence of the putative *Aspergillus oryzae* mismatch repair protein (Ao.MSH2) with the polypeptide sequences of three known mismatch repair proteins from human (msh2-human.p), mouse (msh2-mus.p), and yeast (S.c. msh2).

30

DETAILED DESCRIPTION OF THE INVENTION

A filamentous fungal cell, as described herein, wherein a gene, as described herein, involved in the mismatch repair system has been inactivated.

35

Inactivation of a gene involved in the mismatch repair system:

The NOVEL gene, as described herein, involved in the mismatch repair system may be inactivated by any of the numerous known techniques known to the skilled person.

An embodiment of the invention relates to a filamentous
5 fungal cell as described herein, wherein the gene involved in the mismatch repair is defective.

Numerous methods are known to the skilled person to make a gene defective when the DNA sequence is KNOWN. These methods includes deleting part of the DNA sequence of the gene;
10 introducing frame-shift mutations by deleting or inserting nucleotides; introducing stop codons etc.

A preferred embodiment of the invention relates to a filamentous fungal cell as described herein, wherein the gene involved in the mismatch repair has been inactivated
15 transitorily.

Similarly to above, a number of methods are known to the skilled person for doing this, including insertion of a regulable promoter upstream of the gene or e.g. permanently deleting part of the gene on the chromosome followed by
20 inserting a vector (e.g. a plasmid) into the cell which comprises the gene. The plasmid may then comprise a regulable promoter up-stream of the gene or it may be that the plasmid can be removed from the cell when the mismatch repair system shall be inactivated transitorily and then re-inserted into the cell
25 when the mismatch repair system shall be re-activated.

It is within the skilled persons general knowledge to choose the appropriate strategy for a specific technical purpose.

A preferred way to make a filamentous fungal cell which
30 is capable of transitorily inactivate the mismatch repair system as described herein is first to permanently inactive the mismatch repair gene described herein on the chromosome of the cell followed by inserting a plasmid into the cell which comprises the gene, wherein the plasmid is characterised by
35 that it comprises a suitable replication initiating sequence and a suitable selectable marker.

Preferably the suitable replication initiating sequence is AMA1 (Gems, D., et al. (1991, Gene 98:61-67)).

A more detailed description of suitable replication initiating sequences and suitable selectable markers is provided immediately below and in working example 4 herein is provided an example of this strategy using a plasmid comprising 5 AMA1 as replication initiating sequence and AmdS as selectable marker.

Replication initiating sequences

As used herein, the term "fungal replication initiating 10 sequence" is defined as a nucleic acid sequence which is capable of supporting autonomous replication of an extrachromosomal molecule, e.g., a plasmid or a DNA vector, in a fungal host cell, normally without structural rearrangement of the plasmid or integration into the host cell genome. The replication initiating 15 sequence may be of any origin as long as it is capable of mediating replication initiating activity in a fungal cell. Preferably, the replication initiating sequence is obtained from a filamentous fungal cell, more preferably a strain of *Aspergillus*, *Fusarium* or *Alternaria*, and even more preferably, a 20 strain of *A. nidulans*, *A. oryzae*, *A. niger*, *F. oxysporum* or *Alternaria alternata*.

A replication initiating sequence may be identified by methods well-known in the art. For instance, the sequence may be identified among genomic fragments derived from the organism 25 in question as a sequence capable of sustaining autonomous replication in yeast, (Ballance and Turner, *Gene*, 36 (1985), 321-331), an indication of a capability of autonomous replication in filamentous fungal cells. The replication initiating activity in fungi of a given sequence may also be determined by 30 transforming fungi with contemplated plasmid replicators and selecting for colonies having an irregular morphology, indicating loss of a sectorial plasmid which in turn would lead to lack of growth on selective medium when selecting for a gene found on the plasmid (Gems et al, *Gene*, 98 (1991) 61-67). AMA1 35 was isolated in this way. An alternative way to isolate a replication initiating sequence is to isolate natural occurring

plasmids (e.g. as disclosed by Tsuge et al., Genetics 146 (1997) 111-120 for *Alternaria aternata*).

Examples of replication initiating sequences include, but are not limited to, the ANS1 and AMA1 sequences of *Aspergillus* 5 *nidulans*, e.g., as described, respectively, by Cullen, D., et al. (1987, Nucleic Acids Res. 15:9163-9175) and Gems, D., et al. (1991, Gene 98:61-67).

The term "replication initiating activity" is used herein in its conventional meaning, i.e. to indicate that the sequence 10 is capable of supporting autonomous replication of an extrachromosomal molecule, such as a plasmid or a DNA vector in a fungal cell.

The term "without structural rearrangement of the plasmid" is used herein to mean that no part of the plasmid is deleted 15 or inserted into another part of the plasmid, nor is any host genomic DNA inserted into the plasmid.

Filamentous fungal selective marker

The term "selective pressure" is defined herein as culturing 20 a filamentous fungal cell, containing a DNA vector containing a fungal selective marker gene operably linked to a polynucleotide sequence of interest, in the presence of an effective amount or the absence of an appropriate selective agent. The effective amount of the selective agent is defined herein as an 25 amount sufficient for allowing the selection of cells containing the selection marker from cells which do not contain the selection marker.

In a preferred embodiment, the fungal selective marker is selected from the group of genes which encodes a product capable 30 of providing resistance to biocide or viral toxicity, resistance to heavy metal toxicity, or prototrophy to auxotrophs.

In a more preferred embodiment, the prototrophy is obtained from an enzyme selected from the group of metabolic pathways consisting of nucleotide synthesis, cofactor synthesis, amino 35 acid synthesis, acetamide metabolism, proline metabolism, sulfate metabolism, and nitrate metabolism.

In an even more preferred embodiment, in the methods of the present invention the fungal selective marker is a gene selected from the group consisting of *argB* (ornithine carbamoyl-transferase), *amdS* (acetamidase), *bar* (phosphinothricin acetyl-
5 transferase), *hemA* (5-aminolevulinate synthase), *hemB* (porphobilinogen synthase), *hygB* (hygromycin phosphotransferase), *niaD* (nitrate reductase), *prn* (proline permease), *pyrG* (orotidine-5'-phosphate decarboxylase), *pyroA*, *riboB*, *sC* (sulfate adenyl-transferase), and *trpC* (anthranilate synthase).

10 The fungal cell is cultivated in a suitable medium and under suitable conditions for screening or selecting for transformants harbouring the variant polynucleotide sequence of interest having or encoding the desired characteristic. The cultivation may be performed in accordance with methods well-known in
15 the art for screening of polynucleotide variant libraries.

The filamentous fungal cell

The filamentous fungal cell as described herein includes all filamentous forms of the subdivision Eumycota and Oomycota.
20 The filamentous fungi are characterised by a mycelial wall composed of chitin, cellulose, glucan, chitosan, mannan, and other complex polysaccharides. Vegetative growth is by hyphal elongation and carbon catabolism is obligately aerobic. In contrast, vegetative growth by yeasts such as *Saccharomyces*
25 *cerevisiae* is by budding of a unicellular thallus and carbon catabolism may be fermentative. In a preferred embodiment, the filamentous fungal cell is a cell of a species of, but is not limited to, *Acremonium*, *Aspergillus*, *Fusarium*, *Humicola*, *Mucor*, *Myceliophthora*, *Neurospora*, *Penicillium*, *Scytalidium*,
30 *Thielavia*, *Tolypocladium*, and *Trichoderma*.

Examples of filamentous fungal cells of use in the present invention include an *Aspergillus* cell, an *Acremonium* cell, a *Fusarium* cell, a *Humicola* cell, a *Mucor* cell, a *Myceliophthora* cell, a *Neurospora* cell, a *Penicillium* cell, a *Thielavia* cell,
35 a *Tolypocladium* cell, and a *Trichoderma* cell.

More specifically, the filamentous fungal cell is an *Aspergillus awamori*, *Aspergillus foetidus*, *Aspergillus japonicus*, *Aspergillus nidulans*, *Aspergillus niger*, or *Aspergillus oryzae* cell;

5 a *Fusarium bactridioides*, *Fusarium cerealis*, *Fusarium crookwellense*, *Fusarium culmorum*, *Fusarium graminearum*, *Fusarium graminum*, *Fusarium heterosporum*, *Fusarium negundi*, *Fusarium oxysporum*, *Fusarium reticulatum*, *Fusarium roseum*, *Fusarium sambucinum*, *Fusarium sarcochroum*, *Fusarium*
10 *sporotricioides*, *Fusarium sulphureum*, *Fusarium torulosum*, *Fusarium trichothecioides*, *Fusarium venenatum* cell or a *Fusarium venenatum* cell (Nirenberg sp. nov; a *Humicola insolens* cell or a *Humicola lanuginosa* cell; a *Mucor miehei* cell; a *Myceliophthora thermophila* cell; a *Neurospora crassa*
15 cell; a *Penicillium purpurogenum* cell; a *Thielavia terrestris* cell; or a *Trichoderma harzianum*, *Trichoderma koningii*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, or *Trichoderma viride* cell.

20 A process for preparing a filamentous fungal cell population comprising a DNA library according to the third aspect of the invention:

Placing individually different DNA sequences of interest in a
25 filamentous fungal cell population according to step (a) of the process of the third aspect of the invention:

As state above, the specific experimental way of placing these DNA sequences within a filamentous cell may be done according to any of the many suitable techniques, such as
30 transformation techniques. See the general fungal textbook "Fungal Genetic" (1996, ISBN 0-8247-9544-X) for a further description of such standard techniques.

A practical example may be that single stranded oligonucleotide sequences partially homologous to chromosomal
35 DNA sequence are placed within the cell. See Calissano et al.

(Fungal genetic newsletter 43:15-16 (1995) for further description of this.

Another example may be that duplex DNA sequences comprising mismatches (e.g. comprised within a vector as shown 5 in figure 1) are placed within the cell.

In an preferred embodiment the different DNA sequences of interest is comprised in a plasmid wherein the plasmid is characterised by that it comprises a suitable replication initiating sequence and a suitable selectable marker as 10 described above.

Preferably the suitable replication initiating sequence is AMA1 (Gems, D., et al. (1991, Gene 98:61-67).

Growing the population of step (a) for a period of time 15 allowing an individual DNA sequence of interest in the population to be duplicated at least once under conditions wherein the mismatch repair system gene, as describe herein has been inactivated, according to step (b) of the third aspect of the invention.

20 Growing of the population may be done in any of the numerous suitable known media for growing filamentous fungal cells. It is within the skilled persons general knowledge to choose such a suitable media.

As explained above an individual cell in the population 25 must be allowed to be duplicated at least once under conditions wherein the mismatch repair system gene, as described herein has been inactivated.

Said cells may of course be allowed to be *duplicated more than once* under conditions wherein the mismatch repair system 30 gene has been inactivated.

Since inactivation of the mismatch repair system normally will cause accumulation of mutations on the chromosomal DNA within the cell and thereby maybe make lethal mutations to the cell the actual preferred number of duplication cycles as 35 described above will depend on how fast such potential lethal mutations arise.

It is within the skilled persons general knowledge to determine how many of duplication cycles it preferred.

Due to these potential lethal mutations it is preferred that the mismatch repair system under step (b) has been inactivated transitorily.

After suitable cycles of duplication according to step 5 (b) of the third aspect the transitorily inactivated mismatch repair system the mismatch repair system is then re-activated in order to avoid these lethal mutations in the filamentous fungal cell as such. The strategy for this transitorily inactivation may be any of the strategies described above.

10 Another strategy to limit introduction of mutations on the chromosome, is to transitorily stop the chromosomal replication while replicating the extra-chromosomal element under mismatch repair deficient conditions. This can be achieved by introducing mutations in elements being solely 15 necessary for the chromosomal replication.

A preferred strategy is to use a filamentous fungal cell wherein the gene involved in the mismatch repair system as described herein is permanently inactivated on the chromosome of the cell followed by inserting a plasmid into the cell which 20 comprises the gene, wherein the plasmid is characterised by that it comprises a suitable replication initiating sequence and a suitable selectable marker. See above for a further explanation of this strategy.

Preferably the suitable replication initiating sequence 25 is AMA1 (Gems, D., et al. (1991, Gene 98:61-67).

A further embodiment relates to the process of the third aspect of the invention, wherein the mismatch repair system under step (b) is defective.

In a further embodiment the invention relates to a 30 process as described herein, wherein, under step (b) of the third aspect of the invention, there is an *in vivo* intergenic recombination of partially homologous DNA sequences of interest.

Since the overall concept of the present invention is to 35 provide a process involving inactivation of the mismatch system it is of course preferably that said partially homologous DNA sequences are capable of *in vivo* forming duplex DNA sequences comprising mismatches.

A process for production of a polypeptide of interest comprising the steps of the third aspect of the invention and wherein the DNA sequences of interest encode a polypeptide of interest, according to the fourth aspect of the invention:

Selecting from the resultant population of filamentous fungal cells of step (b) of the third aspect a desired polypeptide of interest, according to step (c) of the fourth aspect.

10 The desired polypeptide of interest may be any polypeptide comprising an desired technical feature, such as improved stability; a desired specific activity; a desired pH optimum; an improved wash performance in a detergent; etc.

15 The specific strategy for selecting this desired polypeptide of interest may be any of the numerous selecting strategies known to the skilled person, such as plate screening assays, micro-titer plate based assays, etc.

20 An embodiment of the invention relates to a process of the fourth aspect of the invention, which further comprises following steps:

25 (d) an optionally step comprising modifying the amino acid sequence of the desired selected polypeptide of interest according to a particularly further specific need;

30 (e) placing the DNA sequence encoding the polypeptide of interest of step (c) of the fourth aspect or the modified polypeptide of interest of step (d) into a filamentous fungal cell which is suitable for large scale production of the polypeptide of interest ;

 (f) cultivating the filamentous fungal cell of step (e) in a fermentor of at least 10000 m3 under conditions permitting expression of the polypeptide of interest; and

35 (g) isolating the polypeptide of interest.

 This embodiment relates to an industrial very relevant process, wherein the selected polypeptide of interest is produced in large scale.

The optionally step (d) relates to a situation wherein e.g. the desired polypeptide of interest is selected in order to e.g. identify a polypeptide with improved wash performance in a detergent according to step (c) of the third aspect of the invention. This polypeptide while having improved wash performance in a detergent may not be sufficiently stable for a commercial product. Accordingly, it may be required to make some further amino acid substitutions in this selected polypeptide, such as e.g. suitable Proline substitutions on order to obtain sufficient stability to commercialising this polypeptide.

A further embodiment relates to a process of the embodiment immediately above, wherein the filamentous fungal cell which is suitable for large-scale production of the polypeptide of interest of step (e) said embodiment is another filamentous fungal cell as compared to the filamentous fungal cell of step (a) of the third aspect of the invention.

This embodiment relates to a situation wherein the filamentous fungal cell used to select the polypeptide of interest is different from the one which is used for large scale production.

A further embodiment relates to a process as described herein, wherein the polypeptide of interest is a polypeptide derived from a filamentous fungal cell.

The term derived from a filamentous fungal cell should be understood in the sense that the information in the amino acid sequence of the polypeptide of interest is derived from a polypeptide obtained from a filamentous fungal cell.

Consequently, it may be a variant of a wild-type filamentous fungal polypeptide and/or may be a polypeptide which is a result of a recombination/shuffling of two or more different filamentous fungal polypeptides.

In an even further embodiment the invention relates to a process as described herein, wherein the polypeptide of interest is an enzyme, such as an amylase, a protease, a cellulase, a lipase, a xylanase; a phospholipase.

EXAMPLES:

Materials

Chemicals used as buffers and substrates were commercial products of at least reagent grade.

5

EXAMPLE 1:

A gel shift assay suitable for determining if a filamentous fungal cell as described herein is inactivated in the mismatch repair system:

10 The principle in this gel shift assay is that cell extracts are prepared of both (a) a filamentous fungal cell wherein the gene, as described herein, involved in the mismatch repair system is inactivated; and (b) the corresponding filamentous fungal cell wherein the gene is NOT inactivated.
15 These extracts are then bound/mixed to oligonucleotides containing the base-pair mismatched G:T; G:A; G:G; A:C, and an extrahelical TG dinucleotide and run on a nondenaturing gel.

If the gel shift assay demonstrates that the control filamentous fungal cell wherein the gene is NOT inactivated
20 comprises any protein(s) which binds to any of above mentioned oligonucleotides and these binding protein(s) is NOT seen in the filamentous fungal cell wherein the gene, as described herein, involved in the mismatch repair system is inactivated then it is a confirmation that the mismatch repair system in
25 the latter is inactivated.

Experimental procedure:

Preparation of cell extracts are performed as described in Nagata et al. (Mol. Gen Genet (1993) 237:251-260; See Materials and Methods).

30 Annealing of oligonucleotides, binding of cell extracts to duplex oligonucleotides containing mismatched, and nondenaturing polyacrylamide gelelectrophoresis are performed essentially as described (Stephenson and Karran; Selective binding to DNA base pair mismatches by proteins from human
35 cells; J. Biol. Chem. 264:2177-21182 (1989)).

However, gelelectrophoresis is performed in TAE buffer rather than in TBE buffer. To obtain duplex oligonucleotides,

the oligonucleotide U is radiolabelled and annealed with any of the unlabelled oligonucleotides L-G.T, L-G.A, L-G.C, L-A.C, L-T.G., or L-HOM. Oligonucleotide sequences are derived from Aquilina et al. Proc. Natl. Acad. Sci. USA 91:8905-8909 (1994).

5

U: 5'-GGGAAGCTGCCAGGCCCCAGTGTTCAGCCTCCTATGCTC-3' (SEQ ID NO 3);

L-G.T: 5'-GAGCATAGGAGGCTGACATTGGGGCCTGGCAGCTTCCC-3' (SEQ ID NO 4) (resulting in a G.T mismatch);

10 L-G.A.: 5'-GAGCATAGGAGGCTGACAATGGGGCCTGGCAGCTTCCC-3' (SEQ ID NO 5) (resulting in a G.A mismatch);

L-G.G.: 5'-GAGCATAGGAGGCTGACAGTGGGGCCTGGCAGCTTCCC-3' (SEQ ID NO 6) (resulting in a G.G mismatch);

15 L-A.C.: 5'-GAGCATAGGAGGCTGACACCGGGGCCTGGCAGCTTCCC-3' (SEQ ID NO 7) (resulting in a A.C mismatch);

L-TG: 5'-GAGCATAGGAGGCTGACACTGTGGGGCCTGGCAGCTTCCC-3' (SEQ ID NO 8) (resulting in an extrahelical TG dinucleotide);

L-HOM: 5'-GAGCATAGGAGGCTGACACCGGGGCCTGGCAGCTTCCC-3' (SEQ ID NO 9) (resulting in a homoduplex).

20

In all assays, a twofold excess of unlabelled homoduplex competitor oligonucleotide is included.

EXAMPLE 2:

25 Cloning of a gene involved in the mismatch repair system of an *Aspergillus oryzae* cell.

The gene cloned as described in this example is shown in SEQ ID NO 1 (DNA sequence) and SEQ ID NO 2 (the translated amino acid sequence).

30 Several sequences of mismatch repair proteins from various organisms are known, only three of these have been utilized in the following: *S. cerevisiae* (M84170), *H. sapiens* (L47580) and mouse (U21011).

The numbers indicated are reference numbers from the
35 public available GenBank database.

Based on the C-terminal homology between known mismatch repair proteins, a set of degenerate primers were designed, aiming at amplification of a partial sequence of the *Aspergillus oryzae* homolog:

5

Pr 117858 (SEQ ID NO 10): P-GGCNCARATHGGNTGYTTYGTNCC

Pr 117859 (SEQ ID NO 11): P-GCCCANGCNARNCCRAANCC

With chromosomal DNA from *A. oryzae* strain JaL142 (WO 96/29391) as template, and above primers the following 50 μ l PWO polymerase based PCR reaction was performed at eight MgSO₄ concentrations (0.5 mM to 4.0 mM, as recommended by the manufacturer; Boehringer M.). 1 mM MgSO₄ was found to be optimal and gave a discrete band of appr. 230 bp as would be expected if no introns were embodied in the sequence.

PCR-cycle profile: [96°C; 2 min - 30 cycles of (94°C; 15s - 50°C; 15s - 72°C; 30s) - 72°C, 7 min - 4°C; hold].

The 230bp PCR fragment was blunt end ligated into filled in BamH1 site of pUC19. pUC19 was BamH1 cleaved in presence of calf intestine alkaline phosphatase, followed by filling in the sticky ends by klenow polymerase and dNTP. Three individual plasmids harbouring the insert were isolated from *E. coli* XL1 transformants of above ligation, and sequenced. Alignment of polypeptides derived by translation of the cloned PCR fragments, revealed a strong homolgy to known mismatch repair protein sequences (see Figure 2).

The underlined sequences of Figure 2 are sequences derived from the consensus PCR primers described above.

The three *Aspergillus* sequences of Figure 2 are equal to the sequence shown in SEQ ID NO 2 from positions 683-758, except from position 685 which in the final cloned sequence is a Thr (T) instead of an Ile (I) as indicated above. This is due to the sequence in above mentioned consensus primers.

The alignment shown in Figure 2 clearly demonstrates that the cloned fragment originates from an *A. oryzae* homologue of a mismatch repair protein.

In order to clone the entire gene, a radiolabeled probe of the cloned fragment was generated by PCR, using 0.5 mg pUC19'msh2'-13 (see above) as template in a 100 ml reaction with Taq polymerase, 30 pmol pUC forward and reverse primers 5 and 0.2 mM of dG-, dC-, dTTP and 0.2mM dATP + ³²P-dATP. The generated radiolabeled probe was liberated from pUC19 sequences by EcoR1- Hind3 digestion followed by gel purification of the resulting 293 bp fragment.

The probe was hybridized to a membrane gridded cosmid library of genomic DNA from *A. oryzae* strain A1560 (the father of JaL142) (WO96/29391). A positive clone was identified on the filter when analyzed in a phosphoimager, and the clone was identified as λ31A2.

The λ31A2 cosmid DNA was propagated and used for southern analysis, using the same radiolabeled primer as above. An approximately 9 Kb Pst fragment, split by BstX (previously found in the cloned PCR fragment) into 5.8 and 3.2 kb fragments both lightening up with the probe, was identified and cloned into Pst cut pUC19, giving a plasmid named pUC19msh2P. The insert was sequentially sequenced, starting with primers pointing out from the previously determined sequence, followed by primers based on the sequences determined in the last run:

```

130740 (SEQ ID NO 12): GCTCGAAACATCCAACATCC
25 130741 (SEQ ID NO 13): GCTGTGAATCACTTGCACC
    131928 (SEQ ID NO 14): CTTTCATAAACTGCGACAAATCATGC
    131929 (SEQ ID NO 15): GGAGGAGCATCTTCGC
    131930 (SEQ ID NO 16): GGAAGTTGAAGACTTTACTTCATCC
    134608 (SEQ ID NO 17): CCAGAACTCGCTAACACC
30 134609 (SEQ ID NO 18): GTGCTTTGCGGACGC
    134610 (SEQ ID NO 19): CAGGACAGTAGGACGC
    135320 (SEQ ID NO 20): CGAGCGATGAACTCTGC
    135321 (SEQ ID NO 21): GCGTTGGTGGATTATCC
    136105 (SEQ ID NO 22): CGTTGCATCTATCATATACC
35 136106 (SEQ ID NO 23): GGTATATGATAGATGCAACGC

```

The 3825 bp sequence hereby determined (SEQ ID NO 1) was translated in the frame previously determined in the PCR

fragment. The resulting protein (SEQ ID NO 2) called Ao.MSH2 was aligned to the protein sequences of known mismatch repair proteins in Figure 3. From the alignment in shown in Figure 3 the cloned and sequenced DNA clearly encompasses the coding
5 sequence for a homolog of yeast, man and mouse mismatch repair proteins, with one intron in the N-terminal part. The position of the intron was deduced by the standard splice rules, and constitutes the only possibility.

10 **EXAMPLE 3:**

Disruption of the gene cloned in example 1 on the chromosome of an *Aspergillus oryzae* cell:

For the disruption experiment the msh2 CDS was deleted from pUC19msh2P (see example 2) by PCR, introducing a NotI site
15 instead. This was done by the primers:

137208 (SEQ ID NO 24): 5' P-CCGCGTCTCCAACAAGATGAATGG

137207 (SEQ ID NO 25): 5' P-CCGCTTTCTCGGGGTCATAGC

20 In a Pwo polymerase based PCR reaction with 2,5 mM MgSO₄ and 50 pg pUC19msh2P (conditions according to the manufacturer):

PCR-cycle profile: [96°C; 2min - 4 cycles of (94°C; 30s -
25 52°C; 30s - 72°C; 3min) - 25 cycles of (94°C; 30s - 59°C; 30s - 72°C; 3min) - 72°C; 10min]

The resulting PCR product of appr. 8.9 Kb was isolated, ligated into pUC19, and transformed into *E. coli* XL1. From the
30 resulting transformants pMsh2Δ was isolated, and the correctness of the new junction and its surroundings verified by sequencing [primer 138149 (SEQ ID NO 26): CCTTTCCACTTTAATCCTAAGC]. (Xl1/pMsh2Δ: Lac3073).

In this construct it the *A.oryzae* pyrG (WO 96/29391) is
35 inserted into the NotI site.

By using this construct the chromosomal gene is deleted in an *Aspergillus oryzae* cell according to standard techniques

known in the art for crossing in such a deleted gene on the chromosome by homologous recombination (Miller, B.L., et al., 1985 Mol. and Cell. Biol. 5:1714-1721).

5 **EXAMPLE 4:**

Construction of a plasmid comprising the mismatch repair gene shown in SEQ ID NO 1, the AMA1 replication initiating sequence; and the AmdS selectable marker:

The plasmid constructed as described below is highly
10 suitable for making a filamentous fungal cell wherein the mismatch repair system may be transitorily inactivated, wherein this plasmid may be inserted into a mismatch disrupted strain of example 3 when the mismatch repair system shall be activated and deleted from the strain when the mismatch repair system
15 shall be inactivated.

Disruption of the mismatch repair gene may cause the accumulation of new chromosomal mutations, thus such a strain might be genetically unstable. Consequently it was decided to perform the chromosomal disruption in a strain where mismatch
20 repair gene was expressed from an extra chromosomal element readily lost when the Δ mismatch repair phenotype was wanted.

The extra-chromosomal element was here a plasmid comprising AMA1 as replication initiating sequence and AmdS as selectable marker.

25 For this purpose the mismatch repair gene (SEQ ID NO 1) was cloned into an autonomously replicating construct harbouring one AMA1 repeat.

From pMT1505 (See example 5 below for description of pMT1505) the following fragments were isolated and ligated
30 together:

5.16 kb NotI-[Hind3]* + 3.515 kb [Sal]*-BamH1 + 757bp
BamH1- NotI

[]* indicates that the site has been filled in by
35 Klenow-polymerase and dNTP

From this ligation reaction pMT1505DHS was isolated (LaC 3212), and the mismatch repair expression cassette was introduced as a BamH1 - Mun1 fragment in the corresponding

sites in pMT1505DHS, resulting in the plasmid pAma-msh2 (LaC 3216).

Aspergillus oryzae JaL250 (see example 5) was transformed AmdS⁺ with pAma-msh2, and the transformants checked for the ability to lose the amdS character when unselected (50% of the transformants), indicating the maintenance of this plasmid as extra chromosomal. (LaC3244 keep on acetamide + uridine).

EXAMPLE 5:

10 Construction of plasmid pMT1505 used in example 4:

Plasmids

pMT1505: constructed as described below in Example 5

pHelp1: contains the *pyrG* gene from *A. oryzae* as a selective marker and the AMA1 sequences which enable autonomous replication in *A. nidulans* as described by Gems, D., et al. (1991. Gene 98: 61-67)

pToC68: as described in EP 0 531 372 (Novo Nordisk A/S)

Strains

20 JaL250: a derivative of *Aspergillus oryzae* A1560 in which the *pyrG* gene has been inactivated, as described in WO 98/01470

DH5a: an *E. coli* host cell purchased from GIBCO BRL (Life Technologies, Inc., Rockville MD)

25

pMT1466 was constructed by inserting an SphI/NarI fragment from pHelp1 into pToC68. pMT1489 was constructed by digesting pMT1466 with SphI and StuI, then religating. pMT1500 was constructed by digesting pMT1489 with AatII and NarI and ligating a linker. pMT1504 was constructed by digesting pMT1500 with NheI and religating.

pMT1505 was constructed by inserting a 2.7 kb XbaI fragment containing the *amdS* encoding gene from *A. nidulans* genomic DNA (Corrick, C.M., et al. 1987, Gene 53:63-71) into pMT1504 which had been cut with NheI.

EXAMPLE 6:**Deletion of part of the *mshII* gene on the chromosome**

The plasmid p418MsHII (from lac3159) is cut with SalI and XhoI and treated with calf-intestinal phosphatase. In this manner part of the *mshII* gene is cut out. The large band (6400 bp) containing the vector and most of the *mshII* gene is isolated from a 1% agarose gel.

The plasmid pJal554 was constructed by ligating a SpeI/SspBI cut fragment (5330 bp) from pSO2 with a Asp718/NheI cut fragment (316 bp) from pSO2. Plasmid pJal554 is cut with SalI and a 2350 bp band which contains the *pyrG* gene is isolated on a 1% agarose gel. The 2350bp band with *pyrG* is ligated with the cut p418MsHII plasmid and transformed into *E. coli*. The right *E. coli* transformant is identified by restriction analysis and a plasmid preparation is made from this transformant.

The plasmid thus prepared is cut with EcoRI in order to linearize the plasmid before it is transformed into for example *Aspergillus oryzae* Jal250. Transformants are selected on mini-10 mal plates.

A transformant where a double crossover event has taken place is identified by making an *Aspergillus* chromosomal DNA prep followed by a PCR screen for full-length *mshII* gene using appropriate primers. A Southern blot is made using chromosomal 15 DNA which is randomly fragmented with appropriate enzymes as well as appropriate probes for the deleted *mshII* fragment (which is not there any longer) as well as a positive control probe.

In order to determine an increased mutation frequency in 20 the strain with an inactivated *mshII* gene, a screen for mutations in the *niaD* gene is made. This is done by growing the parent strain *Aspergillus* Jal250 and the *mshII* inactivated strain on plates.

A spore-suspension is made and aliquotes of spores are 25 plated on chlorate-containing plate as described by Unkles et al. (S. E.Unkles, E.C.Campbell. Y.M.J.T. de Ruite Jacobs, M.

Broekhuisen, J.A. Macro, D.Carrez, R. Contreras, C.A.M.J.J. van den Hondel J.R. Kinghorn. The development of a homologous transformation system for *Aspergillus oryzae* based on nitrate assimilation pathway: A convenient and general selection system for filamentous fungal transformation. Molecular and general genetics V:218 p 99-104 (1989)).

The strain with no expression of the MsHII protein will have a higher rate of niaD mutations (more chlorate resistant clones), than the control strain.

10

EXAMPLE 7:

Making In Vivo antisense msHII RNA to inhibit translation of msHII mRNA using the TAKA promoter to drive transcription of the Anti-sense mRNA

15 Anti-sense RNA expression is a well known way to down regulate expression of any gene in vivo (The design of Antisense RNA , Georg Sczakiel , Antisense and nucleic acid drug development V. 7 P. 439-444 (1997))

A pcr fragment is made using the oligo's:

20 000120j2 (SEQ ID NO 27): TCTGCGAATCGCTTGGATCCCGAACGCGACAACAC,

000120j4 (SEQ ID NO 28): GAGCTCAGATCTCTTAGGTTCTGGACGAGAAGA,

and pUC19msh2P as template. This PCR fragment contains the 5' end of the msHII gene including the presumed part of 5' msHII mRNA. Another PCR fragment is made using the oligo's:

25 000120j3 (SEQ ID NO 29): GTTGTCGCGTTCGGGATCCAAGCGATTTCGCAGAAG,

1298-TAKA (SEQ ID NO 30): GCAAGCGCGCGCAATACATGGTGTGTTTGATCAT,

and pENI1298 as template (PCT DK99/00552).

Both PCR reactions are done using PWO polymerase according to the manufacturers manual (Boehringer-Mannheim).

30 The PCR fragments are purified using the Qiagen PCR purification kit (Qiagen). The two PCR fragments are mixed and a third PCR reaction is done with primer 1298-TAKA and 000120j4. In this manner the two PCR fragments are assembled.

The assembled PCR fragment is cut with BssHII and BglII, 35 and purified from a 1.5 % agarose gel and ligated with pENI1298 which was cut with BssHII and Bgl II (purified from 1 % agarose

gel). The ligation mixture is transformed into *E. coli*. A DNA-prep is made of each of the resulting *E. coli* transformants. The assembled PCR fragment is sequenced to confirm that no unwanted mutations are introduced during the procedure. The correct construct contains the TAKA promoter, which drives the transcription of the *msHII* anti-sense mRNA.

The resulting plasmid is transformed into for example *Aspergillus oryzae* Jal250 along with pENI1298 as control, and transformant are selected on minimal plates. The resulting 10 transformants are isolated on minimal plates and incubated at 37°C until they sporulate.

In order to determine an increased mutation frequency in the strain, where the translation of the *msHII* mRNA is impeded due to *msHII* anti-sense RNA expression, a screen for mutations 15 in the *niaD* gene is made. A spore-suspension is prepared of the control transformants (pENI1298) and of the *msHII* Anti-sense RNA transformants, and equal amounts of spores are plated on to chlorate-containing plate as described by Unkles et al. (S. E.Unkles, E.C.Campbell. Y.M.J.T. de Ruite Jacobs, M. Broekhuizen, J.A. Macro, D.Carrez, R. Contreras, C.A.M.J.J. van den Hondel J.R. Kinghorn. The development of a homologous transformation system for *Aspergillus oryzae* based on nitrate assimilation pathway: A convenient and general selection system for filamentous fungal transformation. Molecular and general genetics 25 ics V:218 p 99-104 (1989)).

The strain with no or low expression of the *MsHII* protein will have a higher rate of *niaD* mutations (more chlorate resistant clones), than the control strain.

CLAIMS

1. A filamentous fungal cell, wherein a gene involved in the mismatch repair system has been inactivated and in which the gene involved in the mismatch repair system comprises:
- 5 (a) a DNA sequence encoding the polypeptide sequence shown in positions 683-758 of SEQ ID NO 2; or
- (b) a DNA sequence encoding a polypeptide sequence which is at least 70% identical to the polypeptide sequence shown in positions 683-758 of SEQ ID NO 2.
- 10
2. A filamentous fungal cell, wherein a gene involved in the mismatch repair system has been inactivated and in which the gene involved in the mismatch repair system comprises:
- (a) a DNA sequence encoding the polypeptide sequence
- 15 shown in positions 1-940 of SEQ ID NO 2; or
- (b) a DNA sequence encoding a polypeptide sequence which is at least 70% identical to the polypeptide sequence shown in positions 1-940 of SEQ ID NO 2.
- 20
3. The filamentous fungal cell of claim 1 or 2, wherein the gene involved in the mismatch repair is defective.
4. The filamentous fungal cell of claim 1 or 2, wherein the gene involved in the mismatch repair has been inactivated
- 25 transitorily.
5. The filamentous fungal cell of any of the preceding claims, wherein the filamentous fungal cell is a strain of *Fusarium* or more preferably *Aspergillus*.
- 30
6. A process for preparing a filamentous fungal cell population wherein individual cells in the population comprise individually different DNA sequences of interest representing a DNA library of interest comprising following steps:
- 35 (a) placing individually different DNA sequences of interest in a filamentous fungal cell population comprising a filamentous fungal cell of claim 1 or 2; and

(b) growing the population of (a) for a period of time allowing an individual DNA sequence of interest in the population to be duplicated at least once under conditions wherein the mismatch repair system gene of claim 1 or 2 has been inactivated.

7. The process of claim 6, wherein the mismatch repair system under step (b) is defective.

8. The process of claim 6, wherein the mismatch repair system under step (b) has been inactivated transitorily.

9. The process of any of claims 6-8, wherein the filamentous fungal cell is a strain of *Fusarium* or more preferably *Aspergillus*.

10. The process of any of claims 6-9, wherein, under step (b) of claim 6, there is an *in vivo* intergenic recombination of partially homologous DNA sequences of interest.

11. A process for production of a polypeptide of interest comprising the steps of claim 6 and wherein the DNA sequences of interest encode a polypeptide of interest and which further comprises following step:

(c) selecting from the resultant population of filamentous fungal cells of step (b) of claim 6 a desired polypeptide of interest.

12. The process of claim 11, which further comprises following steps:

(d) an optionally step comprising modifying the amino acid sequence of the desired selected polypeptide of interest according to a particularly further specific need;

(e) placing the DNA sequence encoding the polypeptide of interest of step (c) of claim 11 or the modified polypeptide of interest of step (d) into a filamentous

fungus cell which is suitable for large scale production of the polypeptide of interest;

(f) cultivating the filamentous fungus cell of step (e) in a fermentor of at least 10000 m³ under conditions permitting expression of the polypeptide of interest; and

(g) isolating the polypeptide of interest.

13. The process of claim 12, wherein the filamentous fungus cell which is suitable for large scale production of the polypeptide of interest of step (e) of claim 12 is another filamentous fungus cell as compared to the filamentous fungus cell of step (a) of claim 6.

14. The process of any of claims 11-13, wherein the polypeptide of interest is a polypeptide derived from a filamentous fungus cell.

15. The process of any of claims 11-14, wherein the polypeptide of interest is an enzyme, such as an amylase, a protease, a cellulase, a lipase, a xylanase; a phospholipase.

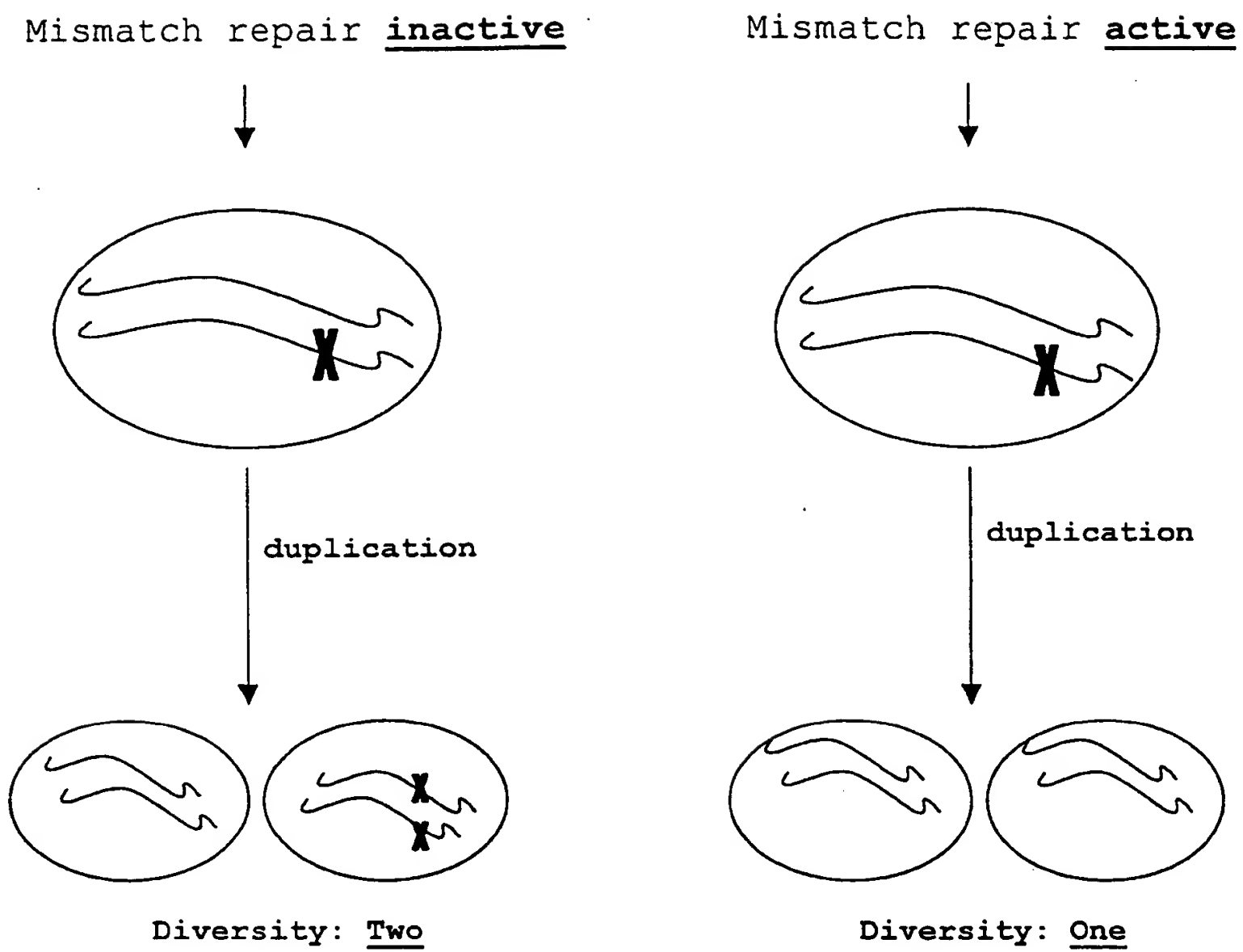


Fig. 1

Contig# 1
'msh2'Ao-coll10
'msh2'Ao-coll13
'msh2'Ao-coll15
msh2-human.p
S.c. msh2

..... ..
AQIGCFVPCTEAELTIFDCILARVGASDSQLKGVSTFMAEMLETSNILKSATSESLIIIDELGRGTSTYDGFGLAW
AQIGCFVPCTEAELTIFDCILARVGASDSQLKGVSTFMAEMLETSNILKSATSESLIIIDELGRGTSTYDGFGLAW
AQIGCFVPCTEAELTIFDCILARVGASDSQLKGVSTFMAEMLETSNILKSATSESLIIIDELGRGTSTYDGFGLAW
AQIGCFVPCESAEVSI VDCILARVGAGDSQLKGVSTFMAEMLETSILRSATKDSLIIIDELGRGTSTYDGFGLAW
AQIGCFVCEEAEIAI VDAILCRVGAGDSQLKGVSTFMVEI LETASILKNASKNSLII VDELGRGTSTYDGFGLAW

70
|

	10	20	30	40	50	60
Contig# 1						
S.c. msh2	o	o	o	o	o	o
Ao.MSH2	MSS-RPELV-D--DEVGIRFYRSLAANSNDETIRVFDRGDWYSAHGAKEAFIARTVYKTTILR					
msh2 mus.p	MAVQPKETLQLEGAAEAGFVRFFEGMPEKPT-TVRLFDRGDFYTAHGEDALLAAREVFKTQGVK					
msh2-human.p	MAVQPKETLQLESAAEVGFVRFFQGMPEKPTT-TVRLFDRGDFYTAHGEDALLAAREVFKTQGVK					

	70	80	90	100	110	120	130
Contig# 1							
S.c. msh2			o		o	o	ooo
Ao.MSH2	NCQLDPVTAKNFHEPTKYVTVSLQVLATLLKCLLDLGYKVEIY-----DKGWKLIKASAP						
msh2 mus.p	N--LGRSDSGGLPS----VTMSVTVFRNFLREALFRLNKRIEW----GSVGTGKGHWKLVKQASP						
msh2-human.p	Y--MGPAGSKTLQS----VVLKMNFESEFVKDLLVRQYRVEVYKNKAGNKASKENEWYLAFAKASP						

	140	150	160	170	180	190
Contig# 1	oo				oo	o
S.c. msh2	GNIEQVNELM----NMNIDSSIIIASLKVQWNSQDGNCCIIGVAFIDTTAYKVGMLDIVDNEVYSNL					
Ao.MSH2	GNLQDVEEELGSLSDSAPIILAVKIS-AKAAEARSVGCFCADASVRELGVSEFLDNDIYSNF					
msh2 mus.p	GNLSQFEDIL--FGNNDMSASVGMGIKMA-VVDGQ-RHVGVGVDSTQRKLGCLCEFPENDQFSNL					
msh2-human.p	GNLSQFEDIL--FGNNDMSASIGVVGKMS-AVDGQ-RQVGVGVDSIQRKLGCLCEFPDNDQFSNL					

	340	350	360	370	380	390
Contig# 1						
S.c. msh2	o		oo	ooo	o oo o oooo	o o oo
	GPQNPFGSNNLAVSGFTSAGNSGKVTSLFQLLNHCKTNAGVRLLNNEWLKKQPLTNIDEINKRHD	LVD				
Ao.MSH2	PRDGSK-SM	-----	-----	SLFGLLNHCKTPVGSRLLAQWLKQPLMDLAEIEKRQQLVE		
msh2 mus.p	SVEDTTGSQ	-----	-----	SLAALLNKCKTAQGQRLVNQWIKQPLMDRNR	IEERLNLVE	
msh2-human.p	SVEDTTGSQ	-----	-----	SLAALLNKCKTPQGQRLVNQWIKQPLMDKNR	IEERLNLVE	

	530	540	550	560	570	580	590
Contig# 1							
S.c. msh2		o o o o o o o o	o o o o o o o o	o o o o o o o o	o o o o o o o o	o o o o o o o o	o o o o o o o o
DEIHSIHLDSAEDLGFDPKKLLKLENHHLHGWCMLRTRNDAKELRKHKKYIELSTVKAGIFFSTKQ							
Ao.MSH2							
HDMGVEHRRVARDLDQDIEKKLFLENHRVHGWCFLRTRNESGCIRNKREYQECSTQKNGVYFTTST							
msh2 mus.p							
KKMQSTLINAAARGLGDPGKQIKLDSSAQFGYFRVTCKEEKVLRNKNKFSTVDIQKNGVKFTNSE							
msh2 - human.p							
KKMQSTLISAARDLGLDPGKQIKLDSSAQFGYFRVTCKEEKVLRNKNKFSTVDIQKNGVKFTNSK							

	670	680	690	700	710	720
Contig# 1	o	o oo	o oo	o	o	o
S.c. msh2	PKLHPMDSERRTHLISSRHPVLEMQDDISFISNDVTLESGKGDFLIITGPNMGGKSTYIRQVGVIS					
Ao.MSH2	PKMHPRGTGNTV-LKEARHPCMEMQDDISFITNDVALVRDESSFLIITGPNMGGKSTYIRQIGVIA					
msh2 mus.p	PVILEKGKGRII-LKASRHACVEVQDEVAFIPNDVHFEKDKQMFHIITGPNMGGKSTYIRQTGVIV					
msh2-human.p	PAILEKGQGRII-LKASRHACVEVQDEIAFIPNDVYFEKDKQMFHIITGPNMGGKSTYIRQTGVIV					

	730	740	750	760	770	780	790
Contig# 1							
S.c. msh2	oooo oooooo oo o o ooooo ooooooooo o ooo oo o	oooo ooooo ooooo ooooo ooooo ooooo	LMAQIGCFVPCESAEIAIVDAILCRVGAGDSQLKGVSTFMVEILETASILKNASKNSLIIVDELGR				
Ao.MSH2	LMAQTGCFVPCTEAELTIFDCILARVGASDSQLKGVSTFMAEMLETSNILKSATSESLIIIDELGR						
msh2 mus.p	LMAQIGCFVPCESAEVSI VDCILARVGAGDSQLKGVSTFMAEMLETSILRSATKDSLIIIDELGR						
msh2-human.p	LMAQIGCFVPCESAEVSI VDCILARVGAGDSQLKGVSTFMAEMLETSILRSATKDSLIIIDELGR						

	800	810	820	830	840	850
Contig# 1						
S.c. msh2	oooooooooooo o o ooooooooo o o oo o					
Ao.MSH2	GTSTYDGFGLAWAIAEHIAASKIGCFALFATHFHELTSEKLPN-VKNMHVVAHIEKNLKEQKHDD					
msh2 mus.p	GTSTYDGFGLAWAISEHIVTEIRCFLFATHFHELTALADRYPKSVKNLHVVAFIGDGTDDSEDK					
msh2-human.p	GTSTYDGFGLAWAISDYIATKIGAFCMFATHFHELTALANQIP-TVNNLHVLTALTEET-----					
	860	870	880	890	900	910
Contig# 1						
S.c. msh2	o oo o o ooooooooo oo o oo oo					
Ao.MSH2	-----EDITLLYKVEPGISDQSFSGIHVAEVVQFPEKIVKMAKRKANELDLDLKTNNEDLK-----KA					
msh2 mus.p	KSKRNQVTLLYRVEPGICDQSFSGIHVAELVRFPEKVVNMARQKAELEEDFTS-SEQDQSSMAID					
msh2-human.p	-----LTMLYQVKKGVCDDQSFSGIHVAELANFPRHVIACAKQKALELEEFQNIQTSLGCDEAEPAA					
	-----LTMLYQVKKGVCDDQSFSGIHVAELANFPKHVIECAKQKALELEEFQYIGESQGYDIMEPAA					

Fig. 3 (Continued)

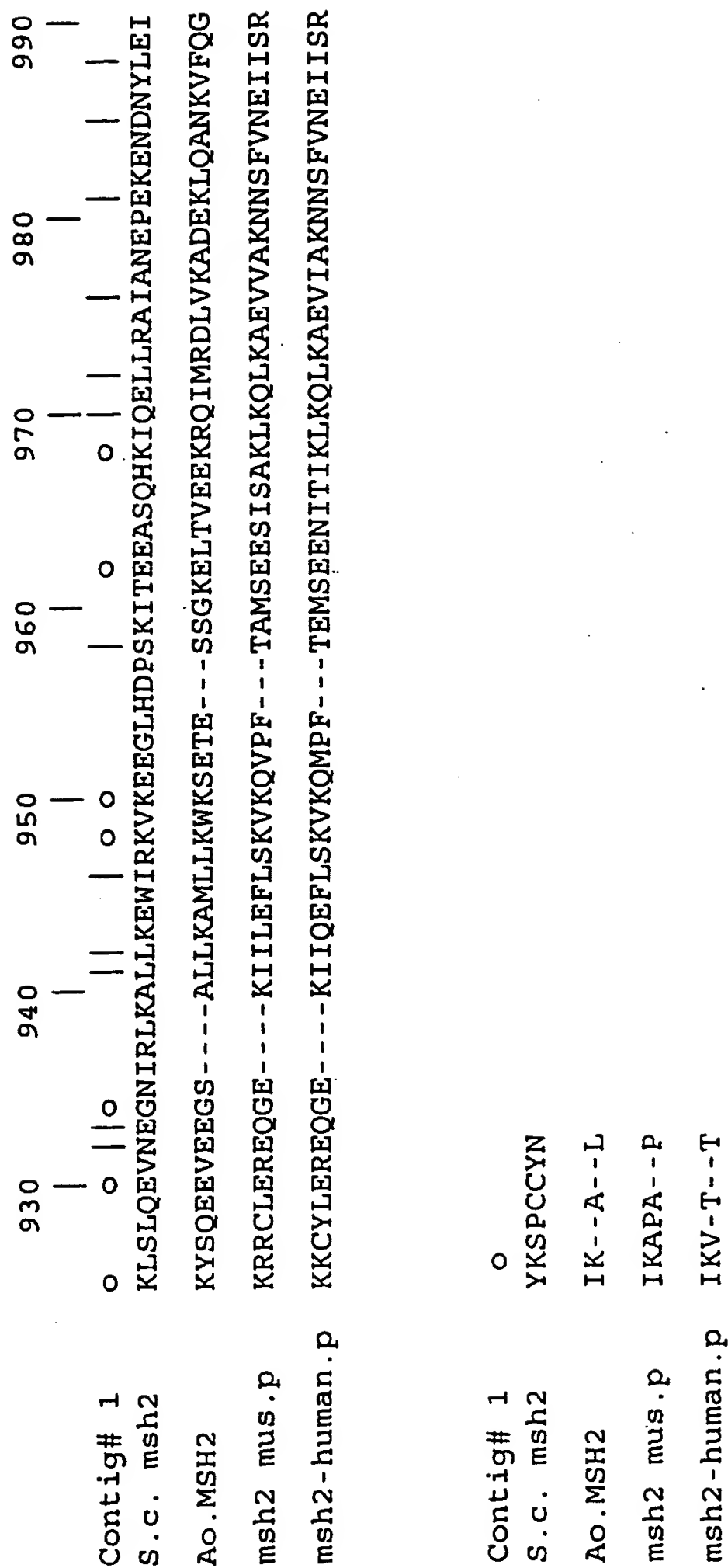


Fig. 3 (Continued)

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Asn Phe Leu Arg Glu Ala Leu Phe Arg Leu Asn Lys Arg Ile Glu Ile	
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Ser	Glu	Gln	Gln	Asp	Gln	Gln	Ser	Ser	Met	Ala	Ile	Asp	Lys	Tyr	Ser
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Glu	Lys	Arg	Gln	Ile	Met	Arg	Asp	Leu	Val	Lys	Ala	Asp	Glu	Lys	Leu
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INTERNATIONAL SEARCH REPORT

International application No.

PCT/DK 00/00063

A. CLASSIFICATION OF SUBJECT MATTER

IPC7: C12N 1/15, C12N 15/10, C12P 9/00 // (C12N 1/15, C12R 1:66), (C12N 1/15, C12R 1:77)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC7: C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 9831837 A1 (MAXYGEN INC.), 23 July 1998 (23.07.98), see page 21, line 31 - page 23, line 20 and page 45, line 12 - page 56, line 21, esp. page 56, line 8 - line 12 --	1-15
A	Database SWISS-PROT, accession no. 013396, Huber D.H.: "DNA Mismatch Repair Protein MSH2. Neurospora crassa." 15 DEC 1998 --	1-15
A	WO 9705268 A1 (SETRATECH), 13 February 1997 (13.02.97), cited in the application --	1-15

☒ Further documents are listed in the continuation of Box C.

☒ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

15 June 2000

Date of mailing of the international search report

16 -06- 2000

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/DK 00/00063

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 9737011 A1 (SETRATECH S.A.R.L.), 9 October 1997 (09.10.97), cited in the application --	1-15
A	WO 9007576 A1 (SETRATECH), 12 July 1990 (12.07.90), cited in the application -- -----	1-15

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/DK 00/00063

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9831837 A1	23/07/98	AU 5920998 A	07/08/98
WO 9705268 A1	13/02/97	AU 4784996 A	26/02/97
		EP 0842289 A	20/05/98
WO 9737011 A1	09/10/97	NONE	
WO 9007576 A1	12/07/90	AT 127519 T	15/09/95
		AU 4834390 A	01/08/90
		CA 2006549 A	26/06/90
		DE 68924174 D,T	14/03/96
		EP 0449923 A,B	09/10/91
		SE 0449923 T3	
		ES 2077058 T	16/11/95
		FR 2641793 A,B	20/07/90
		IE 72469 B	09/04/97
		IL 92856 D	00/00/00
		JP 4503601 T	02/07/92
		US 5912119 A	15/06/99
		US 5965415 A	12/10/99